

Division of Pharmacology Department of Oral Biology

Outline

Our specialty area is “Ca²⁺ signaling”. Ca²⁺ acts as an important intracellular messenger for the regulation of a wide range of biological processes, including secretions, neuronal activities, cell differentiation and migration. We are studying the mechanisms for generating intracellular Ca²⁺ signaling and its functions with a combination of molecular imaging techniques, molecular biology, and gene modification, and trying to elucidate roles of Ca²⁺ signaling in various biological events.

Bio-imaging is a topic of great interest in the bioscience community. Since the isolation of green fluorescent protein (GFP), researchers have reaped an art for creating a tool, which enables the direct visualization of biological functions. We are on the cutting edge of developing biosensors for visualizing inositol 1,4,5 trisphosphate (IP₃), the most important intracellular messenger for triggering Ca²⁺ responses in non-excitabile cells. We use these techniques for studying the regulations of salivary secretions and tooth development.

Saliva secretion is vital for maintenance of oral health and functions. Stimulations of the M₃ muscarinic acetylcholine receptors (mAChRs) resulting in increased flow of salivary secretions through the Ca²⁺ signaling. Most of the studies in this field have been carried out in *in vitro* model systems. Recently, the virus vector-mediated gene delivery allowed us to visualize Ca²⁺ response of salivary glands in live animals with the real time monitoring of salivary secretions and blood flows.

It has been known that amelogenesis imperfecta is caused by the deficiency of store-operated calcium entry (SOCE), the main Ca²⁺ influx mechanisms in non-excitabile cells. These findings suggest important roles of SOCE on amelogenesis. We visualized Ca²⁺ responses in rat dental epithelial cells (SF2) and human dental pulp stem cell (DPSP) using genetically encoded calcium indicators, G-GECO and R-GECO, and examined the role of SOCE on the cell migration and gene expression.

Faculty members (Left→Right)

Professor;

Akihiko TANIMURA, Ph.D.

Associate professor;

Akihiro NEZU, Ph. D.

Assistant professor;

Kaori MURATA, D.D.S., Ph. D.



Postgraduate students (Left→Right)

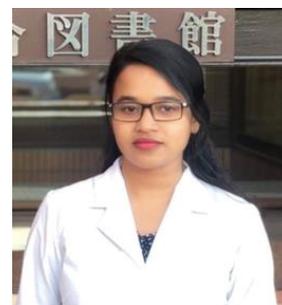
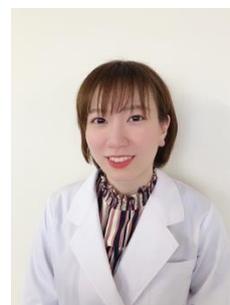
Division of Pharmacology

Azmeree JAHAN, D.D.S.

Narumi ISHIDA, D.D.S.

Tahmina Akter, D.D.S

Rezon Yanuar, D.D.S



Division of Pediatric Dentistry

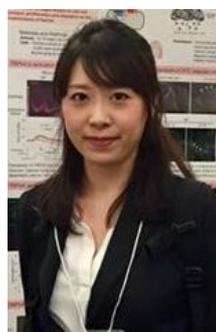
Erika MINOWA, D.D.S.,

Division of Dental Anesthesiology

Kenji GOH, D.D.S.,

Division of Reconstructive Surgery for Oral
and Maxillofacial Region

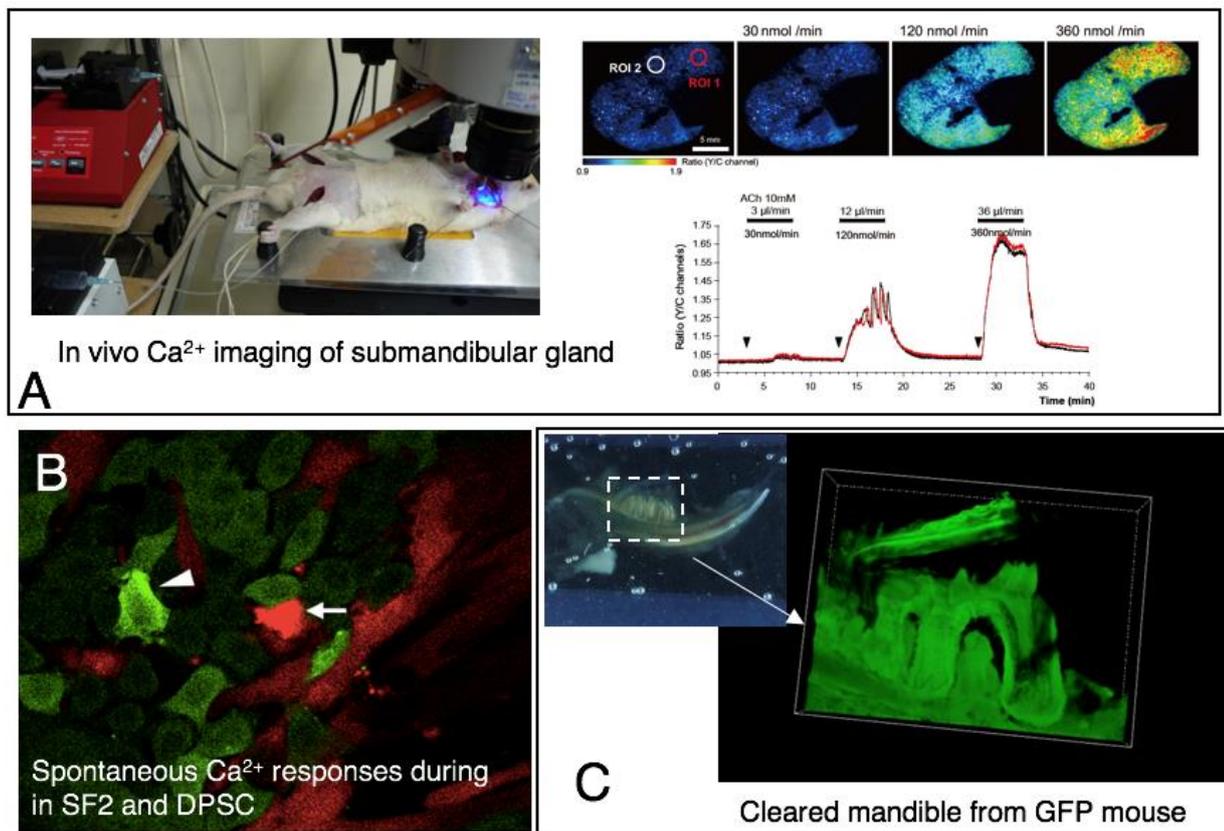
Mari SHIMATANI, D.D.S.,



Undergraduate students are currently doing Research using Tissue Clearing Technology.

Main research in progress

- 1) Development of fluorescent biosensors
- 2) In vivo Ca^{2+} imaging of salivary glands for improving salivary functions (A)
- 3) Role of Ca^{2+} responses for the control of ameloblast differentiation (B)
- 4) Bone and Cancer imaging using Tissue Clearing Technology (C)



Selected publications

- * Takahashi A, Morita T, Murata K, Minowa E, Jahan A, Saito M, Tanimura T. Effects of full-length human amelogenin on the differentiation of dental epithelial cells and osteoblastic cells. *Arch Oral Biol* 107, 104479, 2019.
- * Nezu A, Morita T, Nagai T, Tanimura A. Simultaneous monitoring of Ca^{2+} responses and salivary secretion in live animals reveals a threshold intracellular Ca^{2+} concentration for salivation. *Exp Physiol* 104: 61-69, 2019.
- * Sneyd J, Han JM, Wang L, Chen J, Yang X, Tanimura A, Sanderson MJ, Kirk V, Yule DI. On the dynamical structure of calcium oscillations. *Proc Natl Acad Sci U S A* 114: 1456-1461, 2017.
- * Oura T, Murata K, Morita T, Nezu A, Arisawa M, Shuto S, Tanimura A. Highly Sensitive Measurement of Inositol 1,4,5-Trisphosphate by Using a New Fluorescent Ligand and Ligand Binding Domain Combination. *Chembiochem* 17: 1509-1512, 2016.
- * Murata K, Takahashi A, Morita T, Nezu A, Fukumoto S, Saitoh M, Tanimura A. Effect of 1,25-dihydroxyvitamin D3 on spontaneous calcium responses in rat dental epithelial SF2 cells revealed by long-term imaging. *Biomed Res* 37: 329-334, 2016.
- * Nezu A et al., Partial agonistic effects of pilocarpine on Ca^{2+} responses and salivary secretion in the submandibular glands of live animals. *Experimental Physiology*, 100: 610-651, 2015.
- * Morita T et al., Increase in muscarinic stimulation-induced Ca^{2+} response by adenovirus-mediated Stim1-mKO1 gene transfer to rat submandibular acinar cells in vivo. *Biochem Biophys Res Commun*, 439: 433-7, 2013.
- * Tanimura A et al., A fluorescence-based method for evaluating inositol 1,4,5-trisphosphate receptor ligands: Determination of subtype selectivity and partial agonist effects. *J Biotechnol*, 167: 248-54, 2013.